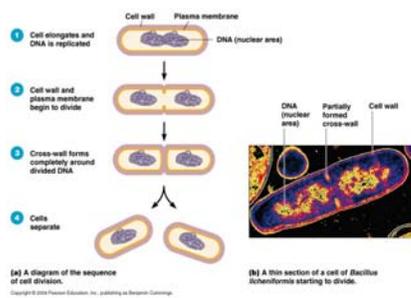


CHAPTER 6 MICROBIAL GROWTH

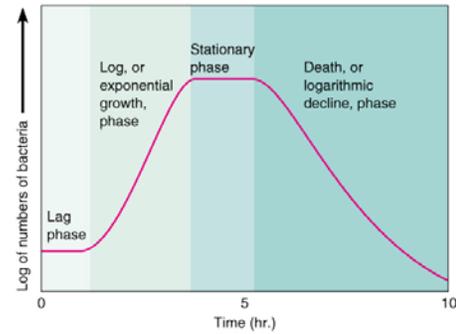
Microbial Growth

- **Growth**= an increase in the number of cells, not an increase in size
- **Generation**=growth by binary fission
- **Generation time**=time it takes for a cell to divide and the population to double; most are 1-3 hours (E.coli: every 20 min.)

Bacteria divide by binary fission



Bacterial growth curve



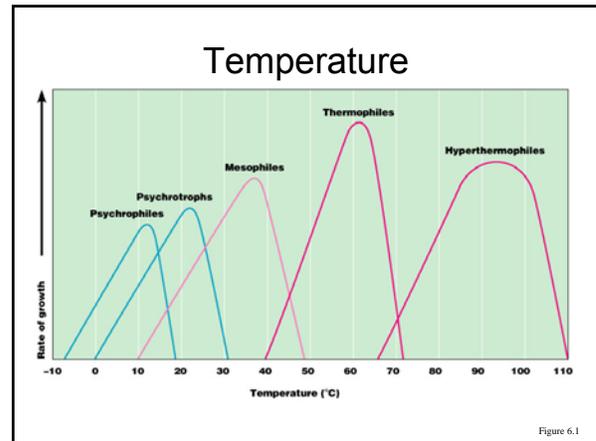
- Chemostat: keeps microbial growth at log stage by draining off used media and replacing with new nutrients.

Requirements for bacterial growth

- Physical requirements
 - Temperature, pH, osmotic pressure, oxygen
- Chemical requirements
 - Carbon, nitrogen, sulfur, and phosphorous
 - Trace elements

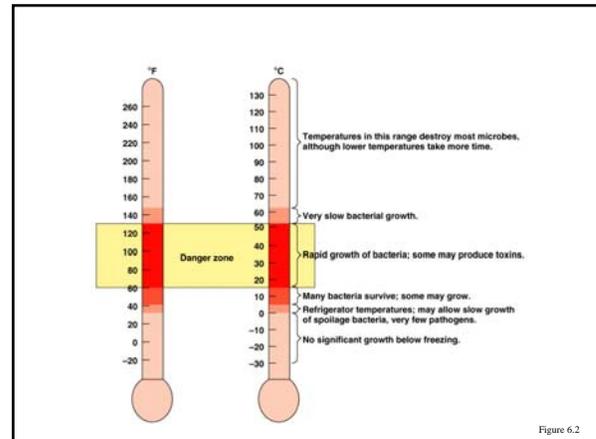
Optimum Growth Temperatures

- Psychrophiles-optimum 15°C
- Psychrotrophs- range from 20-30°C
- Mesophiles- range from 25-40°C
- Thermophiles-range from 50-60°C



PSYCHROTROPHS

- Grow between 0°C and 20-30°C
- Cause food spoilage



- pH
 - Most bacteria grow between pH 6.5 and 7.5
 - Molds and yeasts grow between pH 5 and 6
 - Acidophiles grow in acidic environments

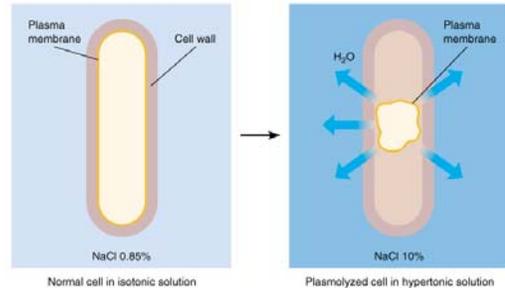
Osmotic Pressure

- Hypotonic
- Isotonic (0.85% NaCl)
- Hypertonic

- Osmotic Pressure

- Hypertonic environments, increase salt or sugar, cause plasmolysis; used to preserve foods
- Extreme or obligate halophiles require high osmotic pressure
- Facultative halophiles tolerate high osmotic pressure

Osmotic environment influences growth



The Requirements for Growth: Chemical Requirements

- Carbon
 - Structural organic molecules, energy source
 - Chemoheterotrophs use organic carbon sources
 - Autotrophs use CO₂

The Requirements for Growth: Chemical

- Nitrogen
 - In amino acids, proteins
 - Most bacteria decompose proteins
 - Some bacteria use NH₄⁺ or NO₃⁻
 - A few bacteria use N₂ in nitrogen fixation
- Sulfur
 - In amino acids, thiamine, biotin
 - Most bacteria decompose proteins
 - Some bacteria use SO₄²⁻ or H₂S
- Phosphorus
 - In DNA, RNA, ATP, and membranes
 - PO₄³⁻ is a source of phosphorus

The Requirements for Growth: Chemical Requirements

- Trace Elements (Iron, Copper, Zinc, etc.)
 - Inorganic elements required in small amounts
 - Usually as enzyme cofactors

The Requirements for Growth: Chemical Requirements

- Oxygen (O₂)

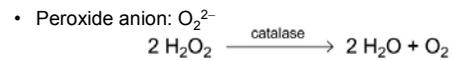
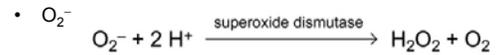
obligate aerobic	Facultative anaerobe	Obligate anaerobe	Aerotolerant anaerobe	Microaerophile

Why can some organisms grow in the presence of oxygen?

- Toxic forms of oxygen need to be neutralized by enzymes
 - Superoxide dismutase
 - Catalase
 - Peroxidase
- If microbe does not produce these enzymes, must have anaerobic conditions.

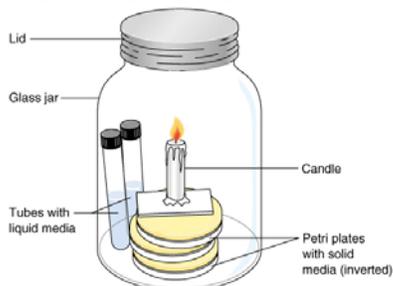
Toxic forms of oxygen

- Singlet oxygen: O_2 boosted to a higher-energy state
- Superoxide free radicals:



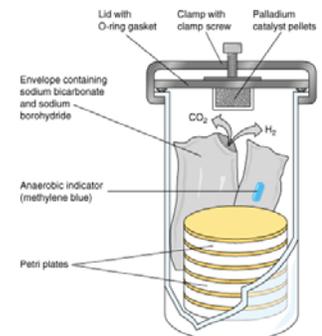
- Hydroxyl radical ($OH\bullet$)

Candle jars increase CO_2 levels for growing capnophiles



(a) Candle jar
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Anaerobic jars eliminate the oxygen for anaerobes to grow.



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The Requirements for Growth: Chemical Requirements

- Organic Growth Factors
 - Organic compounds obtained from the environment
 - Vitamins, amino acids, purines, pyrimidines

Culture Media

- Culture Medium: Nutrients prepared for microbial growth
- Sterile: No living microbes
- Inoculum: Introduction of microbes into medium
- Culture: Microbes growing in/on culture medium

Type	Purpose
Chemically defined	Growth of chemoautotrophs and photoautotrophs, and microbiological assays.
Complex	Growth of most chemoheterotrophic organisms.
Reducing	Growth of obligate anaerobes.
Selective	Suppression of unwanted microbes; encouraging desired microbes.
Differential	Differentiation of colonies of desired microbes from others.
Enrichment	Similar to selective media but designed to increase numbers of desired microbes to detectable levels.

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Agar

- Complex polysaccharide
- Used as solidifying agent for culture media in Petri plates, slants, and deeps
- Generally not metabolized by microbes
- Liquefies at 100°C
- Solidifies ~40°C

Culture Media

- Chemically Defined Media: Exact chemical composition is known
- Complex Media: Extracts and digests of yeasts, meat, or plants
 - Nutrient broth
 - Nutrient agar

Culture Media

- Chemically defined
 - GSA

Constituent	Amount
Glucose	5.0 g
Ammonium phosphate, monobasic (NH ₄ H ₂ PO ₄)	1.0 g
Sodium chloride (NaCl)	5.0 g
Magnesium sulfate (MgSO ₄ · 7H ₂ O)	0.2 g
Potassium phosphate, dibasic (K ₂ HPO ₄)	1.0 g
Water	1 liter

Culture Media

- Complex
 - Nutrient Agar
 - TSA
 - BHI

Constituent	Amount
Peptone (partially digested protein)	5.0 g
Beef extract	3.0 g
Sodium chloride	8.0 g
Agar	15.0 g
Water	1 liter

Anaerobic Culture Methods

- Reducing media
 - Contain chemicals (thioglycollate or oxyrase) that combine O₂
 - Heated to drive off O₂

- Selective media
- Suppress unwanted microbes and encourage desired microbes.

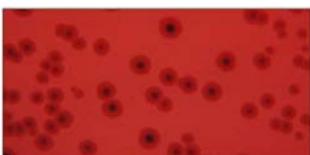


Figure 6.9b, c

Differential Media

- Make it easy to distinguish colonies of different microbes.

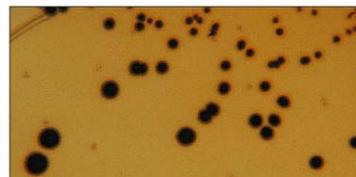
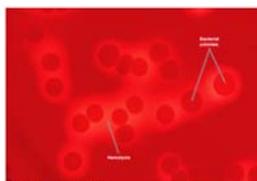


Figure 6.9a

Culture Media

- Selective
- Differential
 - Blood
- Selective and Differential
 - EMB



- Enrichment Media: Encourages growth of desired microbe
- Assume a soil sample contains a few phenol-degrading bacteria and thousands of other bacteria
 - Inoculate phenol-containing culture medium with the soil and incubate
 - Transfer 1 ml to another flask of the phenol medium and incubate
 - Transfer 1 ml to another flask of the phenol medium and incubate
 - Only phenol-metabolizing bacteria will be growing

Transport Media

- Maintains bacterial growth when transporting to laboratory or mailing to state public health lab
- Thayer Martin (Martin-Lewis) Jemec packs for *Neisseria gonorrhoeae*

- A pure culture contains only one species or strain
- A colony is a population of cells arising from a single cell or spore or from a group of attached cells
- A colony is often called a colony-forming unit (CFU)

Streak Plate

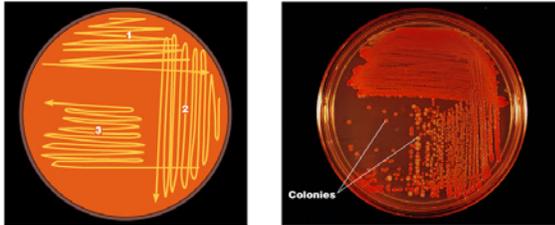
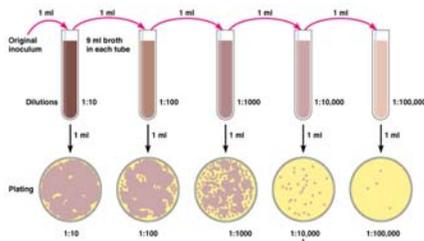


Figure 6.10a, b

Ways to measure bacterial growth

- Plate Count/Viable Cell Count
- Filtration (very small numbers)
- Most Probable Number
- Direct Microscopic Counts
 - Breed count method: Petroff-Hausser counter
- Measure Turbidity; Metabolic activity; Dry weight

Plate counts require dilutions to obtain colonies



Calculation: Number of colonies on plate \times reciprocal of dilution of sample = number of bacteria/ml
 (For example, if 32 colonies are on a plate of $1/10,000$ dilution, then the count is $32 \times 10,000 = 320,000$ per ml in sample.)

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Urine Cultures

- Urine cultures: calibrated loop for .01 or .001 ml of urine
- Cross hatch
- Count colonies:
 - $\geq 10^6$ colonies/ml indicate urinary tract infection

• Filtration

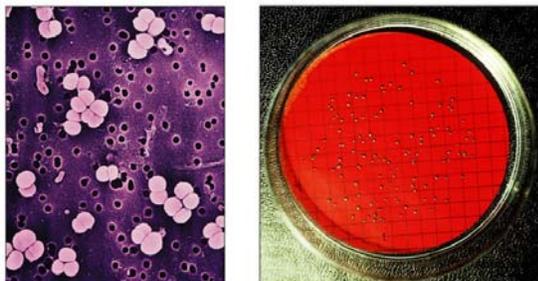


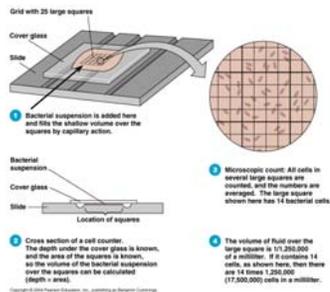
Figure 6.17a, b

- Multiple tube MPN test
- Count positive tubes and compare to statistical MPN table.

Combination of Positives	MPN Index/ 100 ml	95% Confidence Limits	
		Lower	Upper
4-2-0	22	9	56
4-2-1	26	12	65
4-3-0	27	12	67
4-3-1	33	15	77
4-4-0	34	16	80
5-0-0	23	9	86
5-0-1	30	10	110
5-0-2	40	20	140
5-1-0	30	10	120
5-1-1	50	20	150
5-1-2	60	30	180
5-2-0	50	20	170
5-2-1	70	30	210
5-2-2	90	40	250
5-3-0	80	30	250
5-3-1	110	40	300
5-3-2	140	60	360

Figure 6.18b

Direct Counts



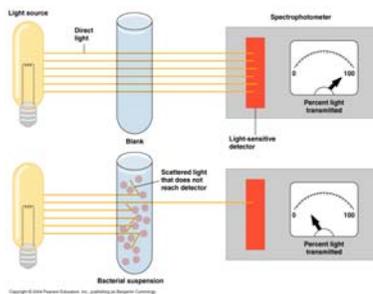
Direct Measurements of Microbial Growth

- Direct Microscopic Count

$$\text{Number of bacteria/ml} = \frac{\text{number of cells counted}}{\text{volume of area counted}}$$

$$\frac{14}{8 \times 10^{-7}} = 17,500,000$$

Turbidity gives a rough estimate



Estimating Bacterial Numbers by Indirect methods

- Metabolic activity
- Dry weight